

University of Montana

## ScholarWorks at University of Montana

---

University of Montana Conference on Undergraduate Research (UMCUR)

---

Apr 28th, 3:00 PM - 4:00 PM

### The Effects of Lipid Structure on Membrane Fluidity

Cynthia Janku

University of Montana, Missoula, [cynthia.janku@umontana.edu](mailto:cynthia.janku@umontana.edu)

Follow this and additional works at: <https://scholarworks.umt.edu/umcur>

**Let us know how access to this document benefits you.**

---

Janku, Cynthia, "The Effects of Lipid Structure on Membrane Fluidity" (2017). *University of Montana Conference on Undergraduate Research (UMCUR)*. 17.

<https://scholarworks.umt.edu/umcur/2017/pmposters/17>

This Poster is brought to you for free and open access by ScholarWorks at University of Montana. It has been accepted for inclusion in University of Montana Conference on Undergraduate Research (UMCUR) by an authorized administrator of ScholarWorks at University of Montana. For more information, please contact [scholarworks@mso.umt.edu](mailto:scholarworks@mso.umt.edu).





# The Effects of Lipid Structure on Membrane Fluidity

Cynthia Janku, Harmen Steele, Dr. Sandy Ross

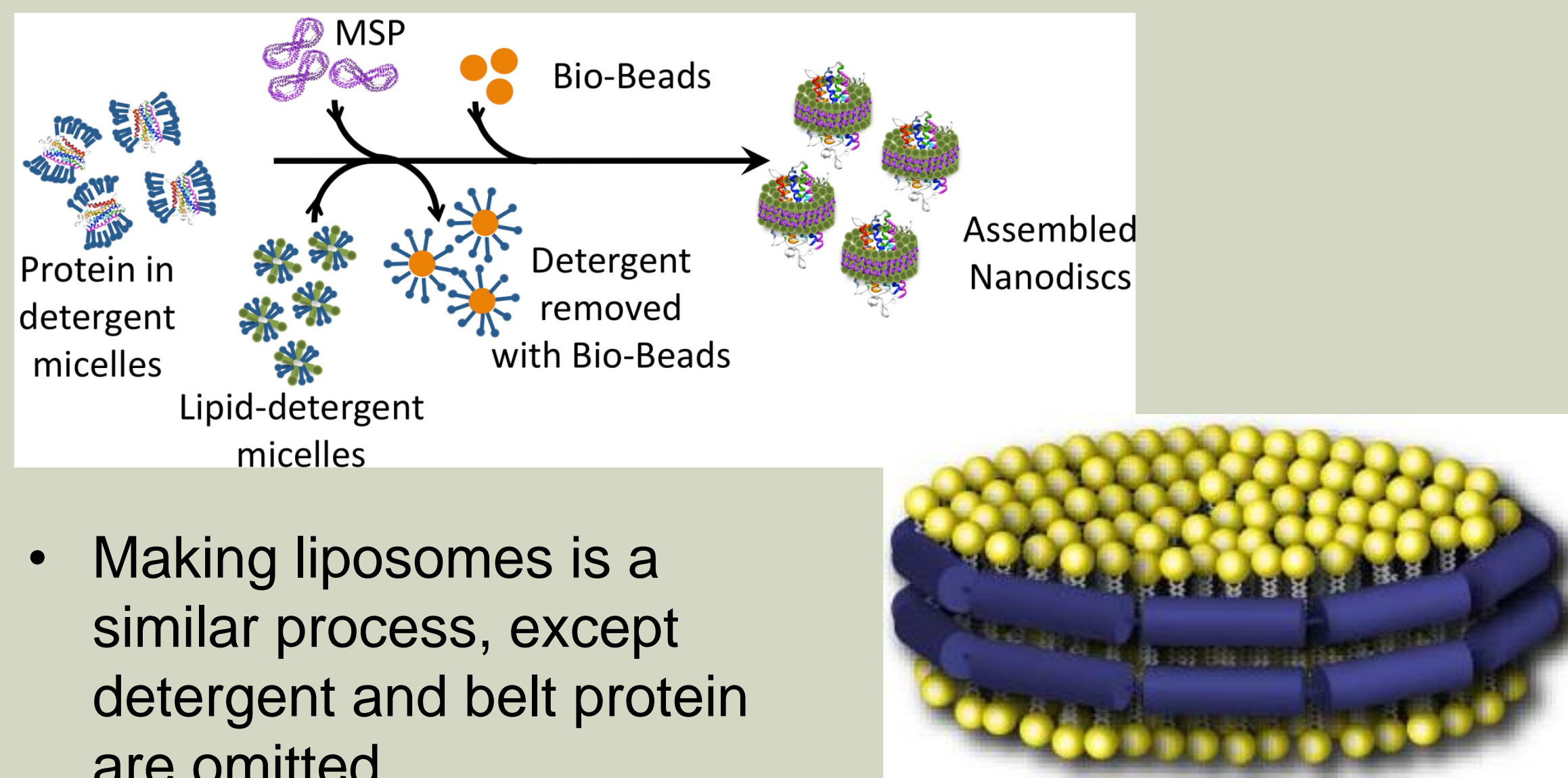
Department of Chemistry and Biochemistry, University of Montana



## Introduction

- Hypothesis: If we excite nanodiscs with different lipid compositions, specifically with differences in saturation, tail length and overall charge, then we would be able to examine the differences in membrane fluidity because the average lifetimes would reflect the relaxation rates.
- Nanodiscs are lipid bilayers surrounded by two identical membrane scaffold proteins, a derivative of apolipoprotein A1.
- Apolipoprotein A1 in the body carries cholesterol and phospholipids through the blood to be excreted by the liver. They are important in fat metabolism.
- We used nanodiscs to study membrane-protein interactions because they are a good model for the lipid-protein interactions in cells. We are using liposomes as a control because they do not have the scaffold protein creating pressure within the macromolecular complex.
- Lifetimes are the average time a fluorophore is in the excited state before releasing a photon and returning to the ground state.
- Intensity decay equation:
$$I(t) = \sum a_n e^{-\frac{t}{\tau_n}}$$

## Making Nanodiscs

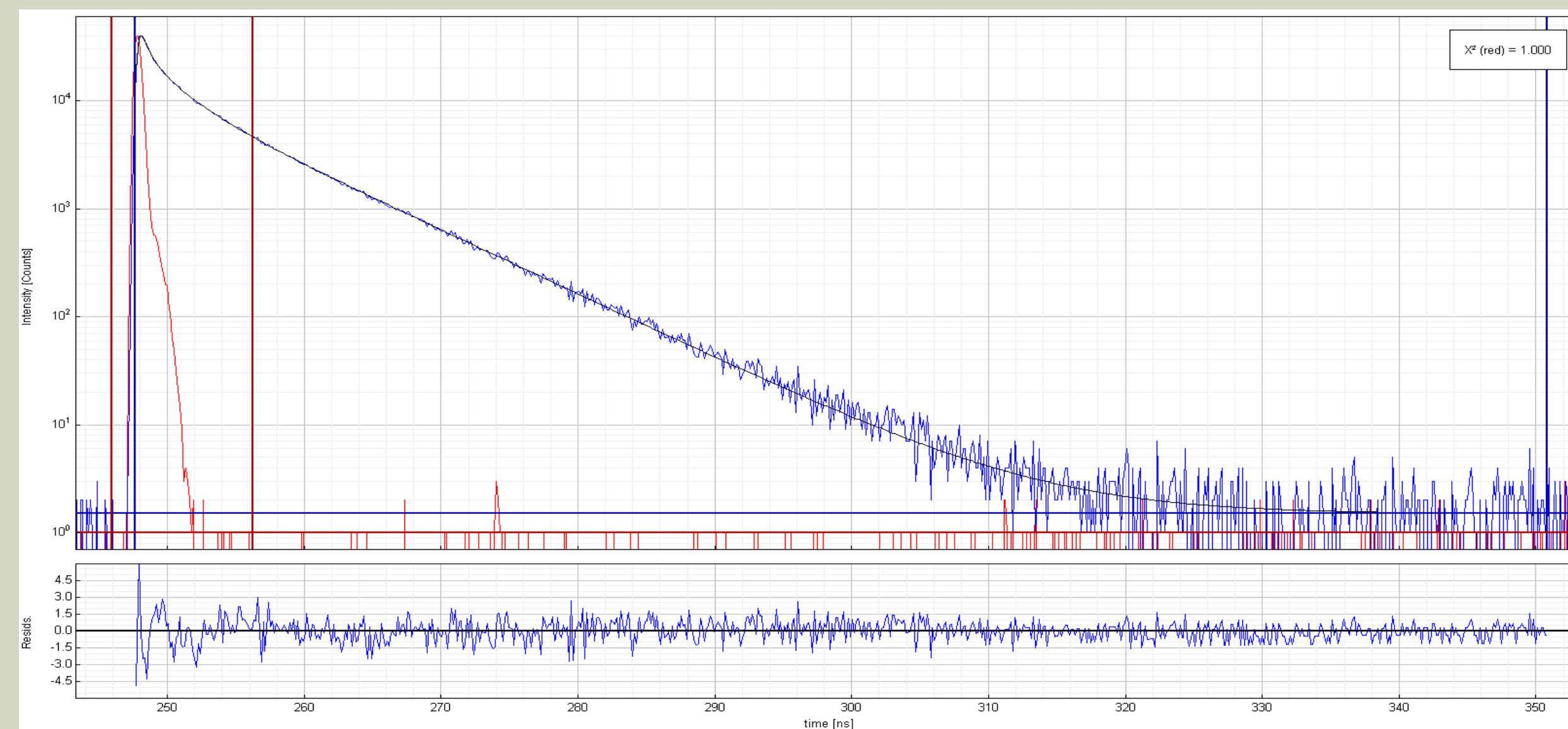


- Making liposomes is a similar process, except detergent and belt protein are omitted.

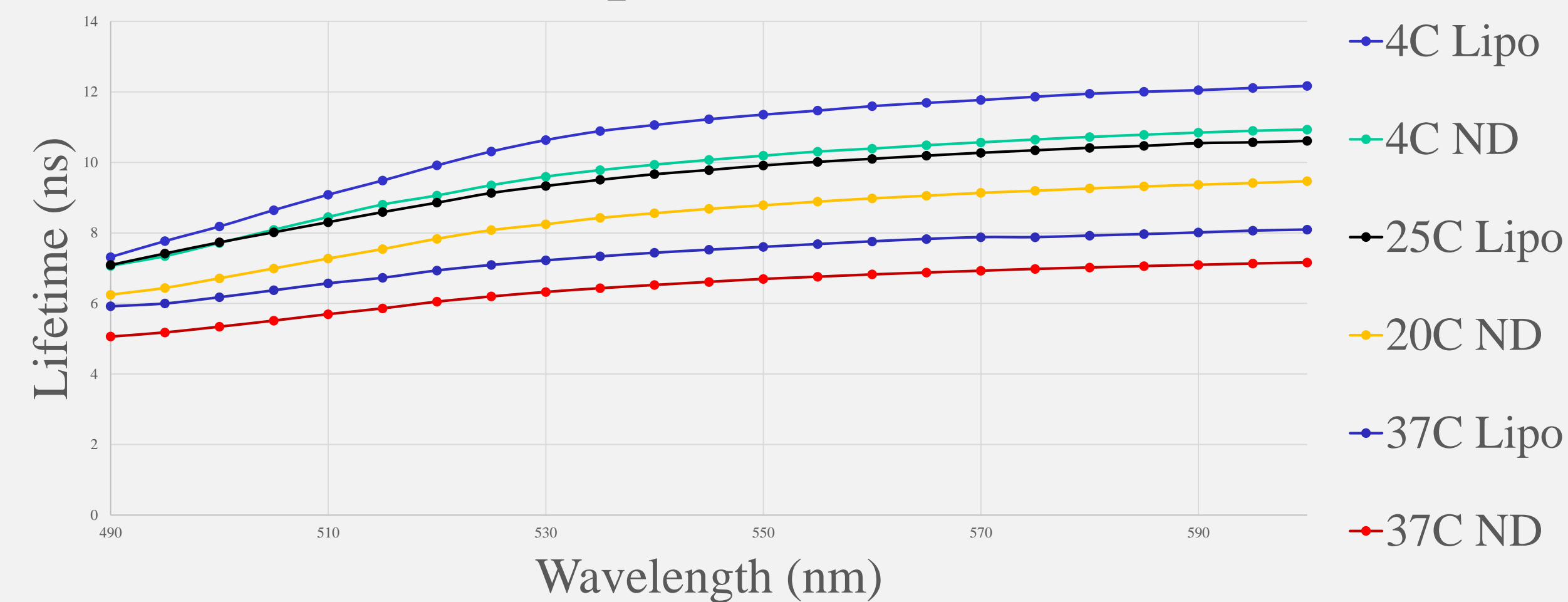
## Lifetimes

- We use FLASC for a sample chamber, TimeHarp to acquire data and FluoFit to analyze the data.
- Excited at vertical angle, the intensity decay is detected at magic angle (54.7°). This eliminates contributions to the intensity decay that could be due to molecular rotation.

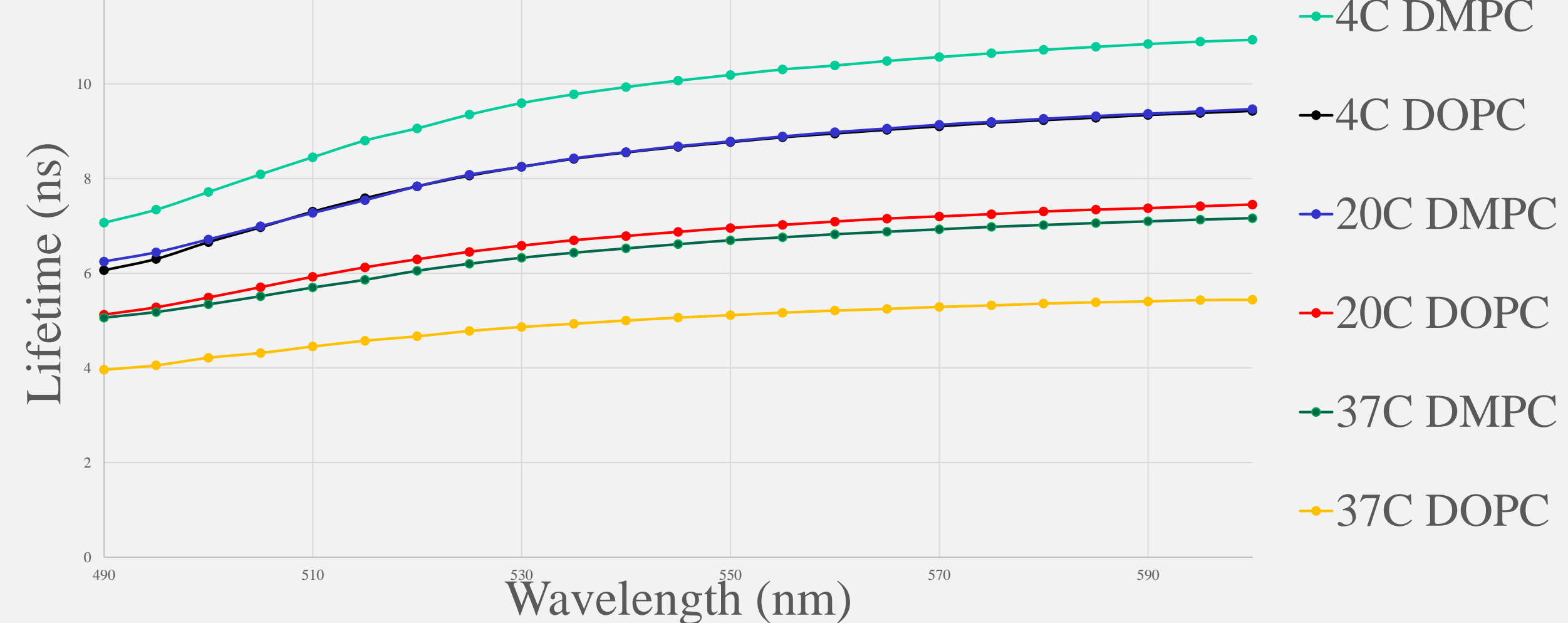
## Intensity Decay Data



## Nanodisc vs Liposome Lifetimes (DMPC)

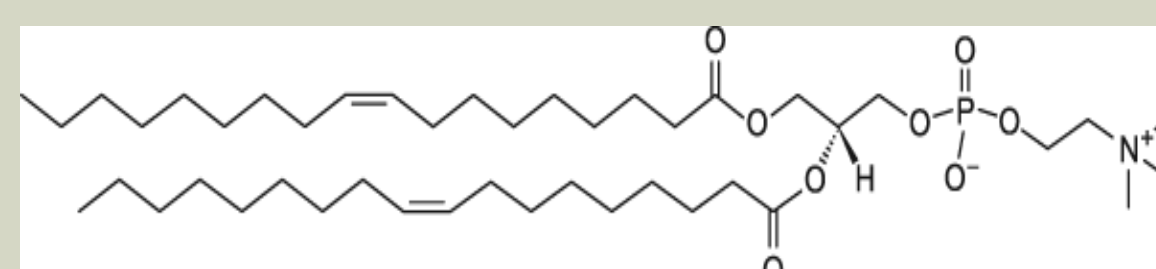


## DOPC vs DMPC Nanodisc Lifetimes

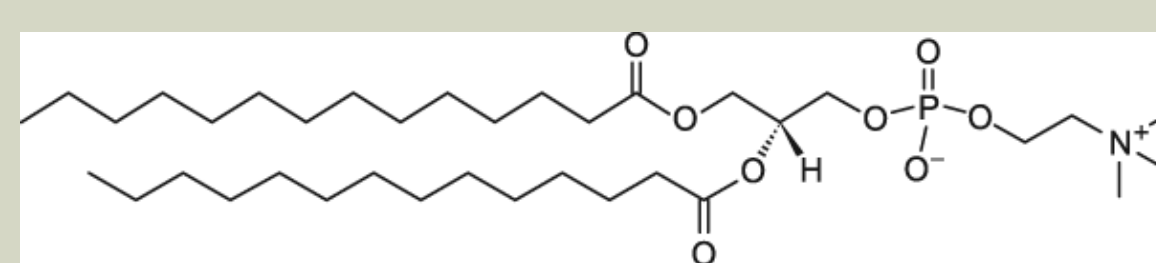


## Lipids Used

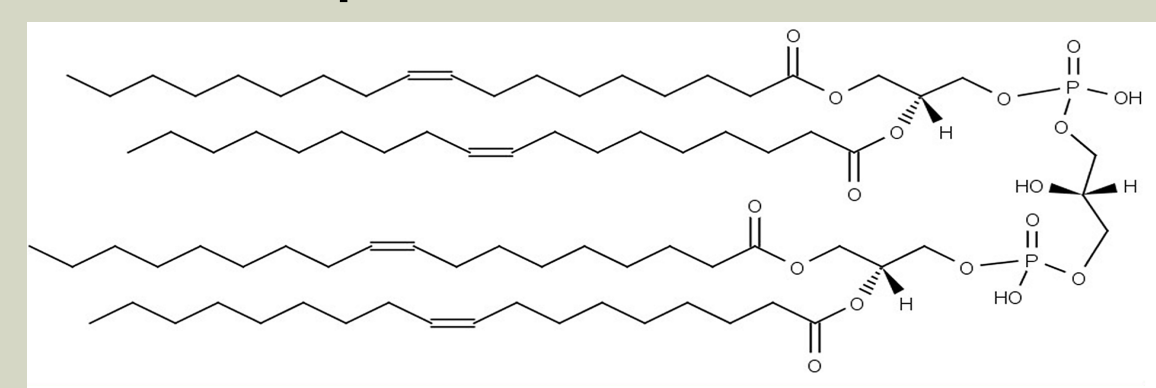
### DOPC



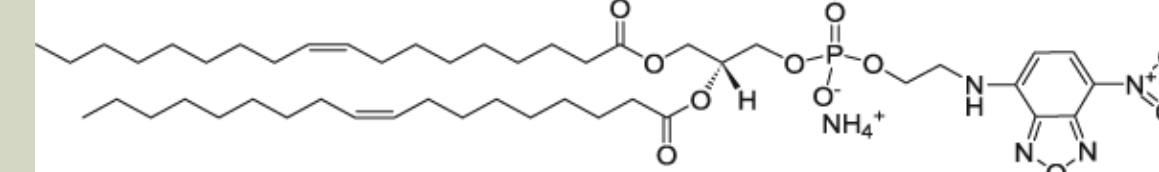
### DMPC



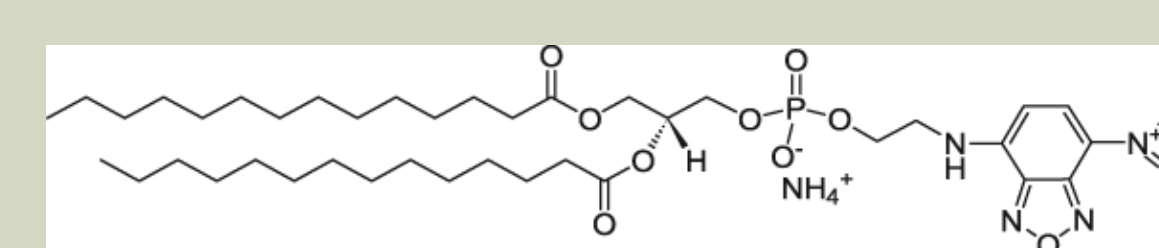
### Cardiolipin:



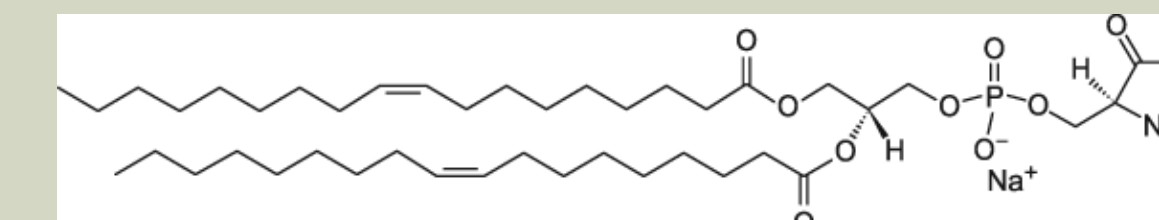
### NBD PE 18:1



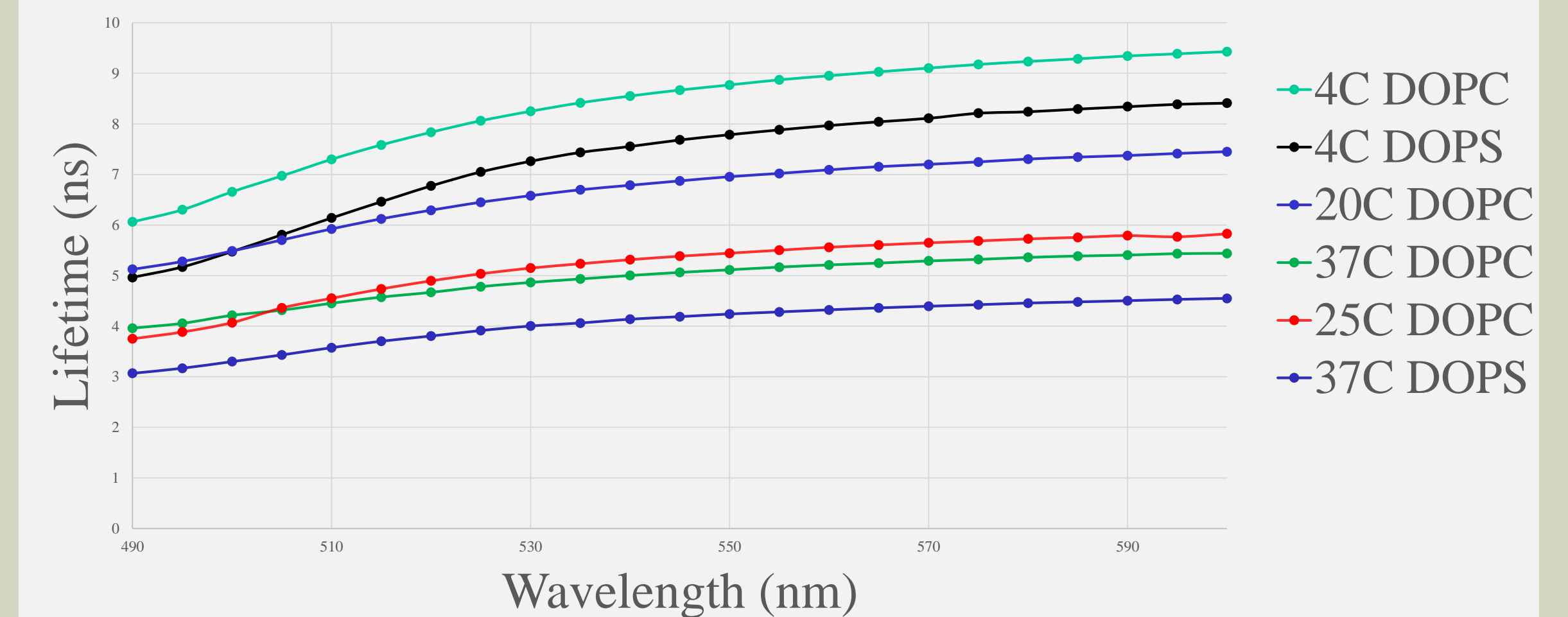
### NBD PE 14:0



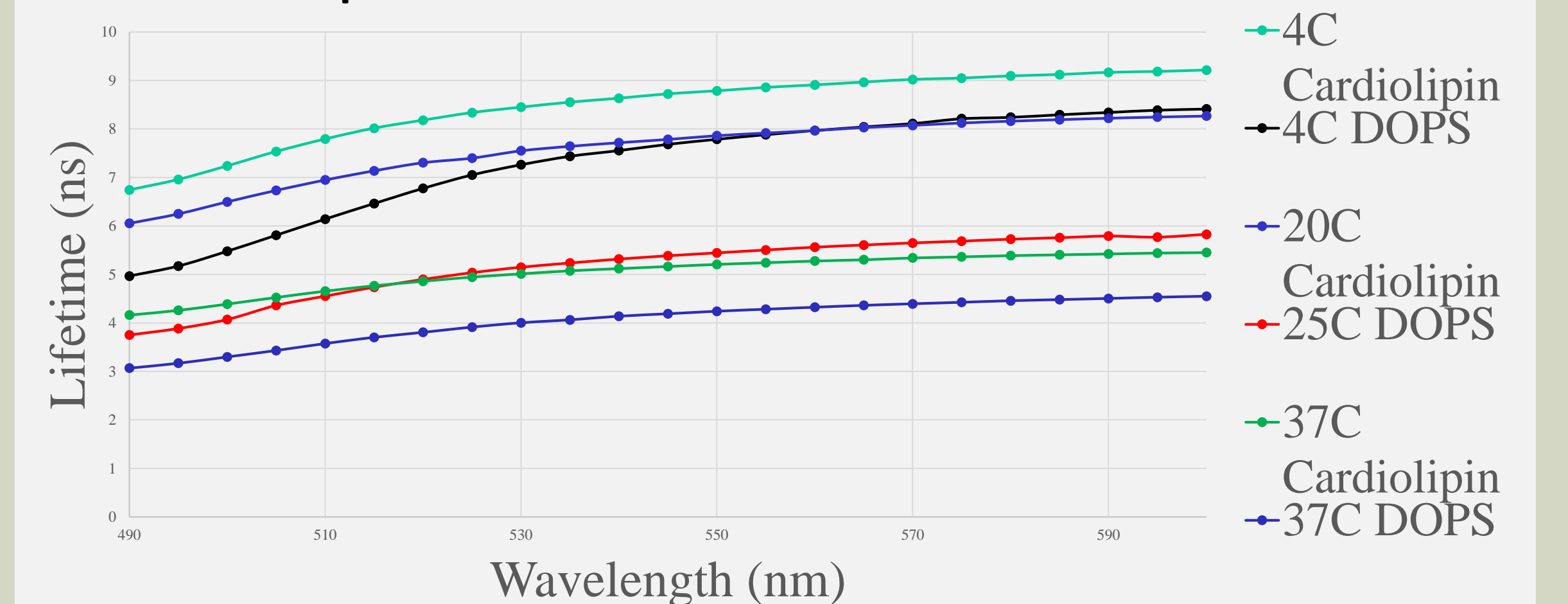
### DOPS



## DOPC vs DOPS Nanodisc Lifetimes



## Cardiolipin vs DOPS Nanodisc Lifetimes



## Results, Conclusions and Implications

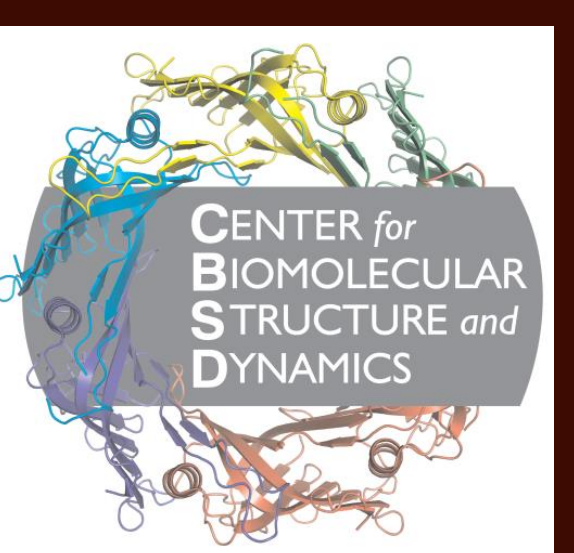
- Lifetimes decrease as temperature increases -nearby interactions increase, thereby increasing the rate of nonradiative decay. In this context, the interactions help the fluorescent lipids release energy, thus reflecting faster relaxation rates at higher temperatures.
- The probes in liposomes have longer lifetimes than in nanodiscs.
- Lifetimes increase as wavelength increases.
- Lipids with unsaturated tails have longer lifetimes.
- The probes in charged environments have shorter lifetimes.

## Future Directions

- Then we will look at TRES, which shows the change in the spectra with respect to wavelength and time, which will reflect solvent relaxation, fluidity, electrostatics and other non-radiative processes.

## Acknowledgments

- Dr. Ross's Lab: Dr. Sandy Ross, Harmen Steele, Kristian Stipe, Sanaa Alabbad, Chelle Terwilliger and William Penny (of Dr. Palmer's Lab)
- Center for Structural and Functional Neuroscience Summer
- Undergraduate Research Fellowship
- NIH CoBRE P20GM103546



## Literature Cited

- Genetics Home Reference, National Institute of Health. 27 July 2015. (ghr.nlm.nih.gov/gene/APOA1)
- www.avantilipids.com
- Picture from www.uni-duesseldorf.de/MathNat/ipb/index.php
- Nivdita, M. et al. Labome, 2013. (www.labome.com/method/Nanodiscs-Membrane-Protein-Research -in-Near-Native-Conditions.html)
- Lakowicz, J.R. et al. Principles of Fluorescence Spectroscopy, 2006, Third Edition.
- Horiba. 2015. (Tech\_Note4\_-\_TRES-DAS.pdf)